Journal of

The Chemical Society,

Chemical Communications

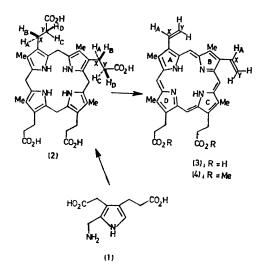
NUMBER 13/1975

Stereochemistry of Biosynthesis of the Vinyl Groups of Protoporphyrin-IX: A Short Synthesis of Porphobilinogen

By ALAN R. BATTERSBY,* EDWARD MCDONALD, HANNS K. W. WURZIGER, and (in part) KEVIN J. JAMES (University Chemical Laboratory, Lensfield Road, Cambridge CB2 1EW)

Summary Porphobilinogen (1), $[^{2}H]$ -labelled in the propionic residue, is synthesised by a short route and is used to establish that both vinyl groups of protoporphyrin-IX are biosynthesised by overall antiperiplanar elimination of a proton and carbon dioxide.

THE biosynthesis of protoporphyrin-IX (3) and thus also of haem involves oxidative conversion of the propionic acid groups on rings A and B of coproporphyrinogen-III (2) into vinyl groups by the enzyme coproporphyrinogenase.¹



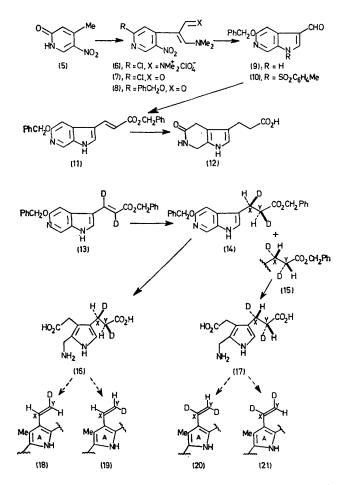
Coproporphyrinogen-III (2) is biosynthesised from four moles of porphobilinogen, PBG (1) by way of uroporphyrinogen-III (2, CH_2CO_2H in place of Me). Earlier studies

from this laboratory² and elsewhere³ showed that the conversion of $(2) \rightarrow (3)$ involved loss of only one hydrogen atom from each of the centres X and retention of both hydrogens at positions Y. Hydrogen removal from the centres X was found to be stereospecific³ with loss of the *pro-S* hydrogens³. We now define the stereochemistry of formation of the vinyl groups.

We developed a synthesis of PBG based on Rapoport's azaindole approach⁴ but new chemistry allowed considerable shortening and more than doubled the yield.

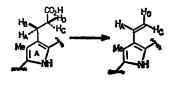
The pyridone⁵ (5) with POCl₃-dimethylformamide (DMF)⁶ in refluxing chloroform (24 h) gave the salt (6). 90% yield, m.p. 233° (decomp.), which was hydrolysed by sodium hydroxide in aqueous acetone to the aldehyde (7) m.p. 189—190° and then (7) with sodium benzyloxide in benzyl alcohol gave the ether (8), 80% yield from (6), m.p. 128—129°. Reduction of (8) with zinc dust and aqueous acetic acid yielded the azaindole (9), 60% yield, m.p. 194—195° which reacted with monobenzyl malonate in dry pyridine-piperidine to form the acrylate (11), 98% yield, m.p. 175—177°. Palladium and hydrogen then cleaved the benzyl groups, saturated the double bond and reduced the pyridone to give PBG lactam (12), 84% yield, characterised as its methyl ester, m.p. 245—247°. The yield of lactam (12) from the pyridone (5) was 35% overall.

The N-tosyl aldehyde (10) was deuteriated by the morpholinonitrile method' and cleavage with NaOD in D_2O -tetrahydrofuran gave, after exchange of >ND against water, the deuteriated aldehyde (9, CDO in place of CHO; no CHO detectable by n.m.r.). This condensed with deuteriated monobenzyl malonate to give the [${}^{2}H_{2}$]-acrylate (13) which was reduced by diimide to form the racemate (14) + (15). In this product, the relative configuration at centres X and Y has been fixed by the established sym-stereospecificity of



diimide. Hydrogenation gave the corresponding PBG lactams and hydrolysis then yielded labelled PBG as a racemate (16) + (17). This was converted by our preparative cell-free system from Euglena gracilis⁸ into protoporphyrin-IX isolated as its ester.

The ¹H n.m.r. signals from H_A of each vinyl group of unlabelled protoporphyrin-IX dimethyl ester (4) appear as a double doublet centred at ca. τ 2.8 (J_{trans} 18 Hz, J_{cis} 11 Hz) which can overlap, but at a suitable concentration eight separate signals can be observed corresponding to the two hydrogens H_A . In the enzymic conversion of labelled PBG (16) + (17) into protoporphyrin-IX, the (R,R)enantiomer (17) will lead either to arrangement (20) or to (21) and neither of these can give rise to a signal at $\tau 2.8$ (no ¹H at X centres). In contrast, the (S,S)-enantiomer (16) by elimination of the forward hydrogen (D in this case) antiperiplanar with the carboxyl group would lead to vinyl groups as in (18) whereas a synperiplanar process would form vinyl groups as in (19). The ¹H n.m.r. spectrum of the labelled protoporphyrin-IX dimethyl ester showed two slightly broadened doublets both with the trans coupling (J 18 Hz) for the two H_A hydrogens and so it was established that both vinyl groups had trans oriented hydrogen atoms (18). The ¹H n.m.r. signals from the centres Y will be analysed in full later; the signal pattern confirmed the observations for the centres X. Thus, biosynthesis of the two vinyl groups of protoporphyrin IX occurs by an overall antiperiplanar elimination of a proton and carbon dioxide (Scheme).



SCHEME

We thank Drs. D. H. Grayson and E. Hunt for their help and the British Council, the Nuffield Foundation and S.R.C. for financial support.

(Received, 24th March 1974; Com. 355.)

¹S. Sano and S. Granick, J. Biol. Chem., 1961, 236, 1173; A. M. del C. Batlle, A. Benson, and C. Rimington, Biochem. J., 1965, 97, 731 and refs. therein.

- A. R. Battersby, J. Baldas, J. Collins, D. H. Grayson, K. J. James, and E. McDonald, J.C.S. Chem. Comm., 1972, 1265.
- ¹ Z. Zaman, M. Abboud, and M. Akhtar, J.C.S. Chem. Comm., 1972, 1263.
 ⁴ B. Frydman, S. Reil, M. E. Despuy, and H. Rapoport, J. Amer. Chem. Soc., 1969, 91, 2338.
 ⁵ D. M. Besly and A. A. Goldberg, J. Chem. Soc., 1954, 2448.
 ⁶ Z. M. Guld G. M. Goldberg, J. Chem. Soc., 1954, 2448.
- ⁶Z. Arnold, Coll. Czech. Chem. Comm., 1963, 28, 863; B. A. J. Clark, J. Parrick, P. J. West, and A. H. Kelly, J. Chem. Soc. (C), 1970, 498. ⁷ D. J. Bennett, G. W. Kirby, and V. A. Moss, J. Chem. Soc. (C), 1970, 2049.

 - ⁸ Cf. E. F. Carell and J. S. Kahn, Arch. Biochem. Biophys., 1964, 108, 1.